

麻楝枝干的化学成分及其 α -葡萄糖苷酶抑制活性研究

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摘要: 为了解麻楝(*Chukrasia tabularis* A. Juss)中的生物活性成分, 采用柱色谱技术从其枝干乙醇提取物中分离得到 10 个化合物, 分别鉴定为: 3-hydroxy-1-(4-hydroxy-3,5-dimethoxyphenyl)propan-1-one (**1**)、6-hydroxy-1,3,5,7-tetramethoxy-9-xanthen-9-one (**2**)、2,6,2',6'-tetramethoxy-4,4'-bis(2,3-epoxy-1-hydroxypropyl)biphenyl (**3**)、cleomiscosin D (**4**)、chuktubularin A (**5**)、chuktubularin B (**6**)、chubularisin H (**7**)、chubularisin I (**8**)、tabularisin A (**9**)和 tabularisin B (**10**), 其中化合物 **1~4** 为首次从麻楝属中分离得到。对体外 α -葡萄糖苷酶的抑制活性进行了测定, 结果表明化合物 **1**、**2**、**6**、**7** 和 **9** 对 α -葡萄糖苷酶均具有较好的抑制活性。

关键词: 麻楝; 枝干; 化学成分; α -葡萄糖苷酶抑制活性

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Studies on the Chemical Constituents from the Stems of *Chukrasia tabularis* and Their α -Glucosidase Inhibitory Activity

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Abstract: In order to find the bioactive components from the stems of *Chukrasia tabularis*, ten compounds were isolated from its EtOH extract by using chromatographic techniques. On the basis of spectral data, their structures were identified as 3-hydroxy-1-(4-hydroxy-3,5-dimethoxyphenyl)propan-1-one (**1**), 6-hydroxy-1,3,5,7-tetramethoxy-9H-xanthen-9-one (**2**), 2,6,2',6'-tetramethoxy-4,4'-bis(2,3-epoxy-1-hydroxypropyl) biphenyl (**3**), cleomiscosin D (**4**), chuktubularin A (**5**), chuktubularin B (**6**), chubularisin H (**7**), chubularisin I (**8**), tabularisin A (**9**), and tabularisin B (**10**). Compounds **1~4** were isolated from the genus *Chukrasia* for the first time. Furthermore, compounds **1**, **2**, **6**, **7** and **9** exhibited inhibitory activity against α -glucosidase *in vitro*.

Key words: *Chukrasia tabularis*; Stem; Chemical constituent; α -Glucosidase inhibitory activity

麻楝 (*Chukrasia tabularis* A. Juss) 为楝科 (Meliaceae) 麻楝属植物, 该属为单种属, 仅包括麻楝原种及其变种毛麻楝 (*C. tabularis* var. *velutina*)^[1], 其主要分布于印度、缅甸、斯里兰卡、中南半岛、

马来半岛等地, 我国云南、广东、海南、广西、西藏等地均有分布。据《中华本草》记载, 麻楝的根皮是我国传统中药材, 具有疏风清热的功效, 主要用于治疗感冒发热。麻楝的主要化学成分为柠檬苦

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素、香豆素、黄酮和挥发油等,其中柠檬苦素是麻楝的特征性化学成分^[2]。据报道,麻楝中的柠檬苦素类化合物具有抗炎^[3-4]、钾离子通道阻断^[5-6]、昆虫拒食^[7-8]、抗肿瘤^[9]等多种药理活性。前期活性筛选中发现麻楝乙醇提取物对 α -葡萄糖苷酶具有良好的抑制活性,为了寻找其中具有 α -葡萄糖苷酶抑制活性的成分,我们对麻楝枝干乙酸乙酯部分的化学成分进行了研究,从中分离鉴定了10个单体化合物,其中柠檬苦素类化合物6个。本文报道了从麻楝中分离得到的10个化合物,并测定他们对 α -葡萄糖苷酶的抑制活性。

1 材料和方法

1.1 材料

麻楝枝干于2014年7月采集于海南省海口市,经中国热带农业科学院热带生物技术研究所刘寿柏博士鉴定为麻楝(*Chukrasia tabularis* A. Juss),凭证标本(CTHK201407)存放于中国热带农业科学院热带生物技术研究所。

1.2 仪器和试剂

化合物分离采用青岛海洋化工厂的薄层色谱硅胶板(GF₂₅₄)和柱色谱硅胶(200~300和60~80目);Merck公司的Sephadex LH-20和RP-18填料。旋光度测定采用Autopol III旋光仪;质谱测定采用Autospec-3000质谱仪;核磁共振采用瑞士Bruker公司的Bruker AV-500型超导核磁仪(TMS内标);美国宝特公司ELX-800酶标仪,超净工作台为上海博讯实业有限公司医疗设备厂产品; α -葡萄糖苷酶(α -Glucosidase, EC 3.2.1.2)、4-硝基苯酚- α -D-吡喃葡萄糖苷(4-Nitrophenyl- α -D-glucopyranoside, PNPG)、阿卡波糖(Acarbose)均购自Sigma公司。

1.3 提取和分离

麻楝枝干(110 kg)晒干后粉碎,用95%乙醇冷浸提取3次,室温,每次7 d;过滤,合并滤液后经真空减压浓缩得粗浸膏,将其分散于水中成悬浊液,依次用石油醚、乙酸乙酯、正丁醇萃取,分别得石油醚萃取物30 g、乙酸乙酯萃取物1700 g、正丁醇萃取物800 g。乙酸乙酯萃取物(1700 g)采用硅胶柱色谱,以石油醚-乙酸乙酯(1:0~0:1)梯度洗脱,分段收集得到18个流分(Fr.1~Fr.18)。Fr.17(120 g)继续采用硅胶(硅胶H)减压柱色谱,以氯仿-甲醇(1:0~0:1)

梯度洗脱,分段收集得到18个流分(Fr.17-1~Fr.17-18)。Fr.17-1(7.0 g)经加压ODS(甲醇-水3:7~1:0)梯度洗脱,得21个流分(Fr.17-1-1~Fr.17-1-21)。经反复Sephadex LH-20(氯仿-甲醇1:1)柱色谱以及加压硅胶柱色谱得到化合物**1**(1.5 mg)、**2**(1.8 mg)、**3**(20.0 mg)、**4**(7.0 mg)、**5**(5.2 mg)和**6**(9.0 mg);Fr.15(268 g)采用硅胶(硅胶H)减压柱色谱,以氯仿-乙酸乙酯(1:0~0:1)梯度洗脱,分段收集得到8个流分(Fr.15-1~Fr.15-8)。Fr.15-2(36.8 g)经MCI(甲醇-水7:1~1:0)梯度洗脱,得到6个流分(Fr.15-2-1~Fr.15-2-6)。Fr.15-2-3(3.5 g)经加压ODS(甲醇-水3:1~1:0)梯度洗脱,获得20个流分(Fr.15-2-3-1~Fr.15-2-3-20)。经反复Sephadex LH-20(氯仿-甲醇1:1)色谱及硅胶柱色谱得到化合物**7**(5.4 mg)、**8**(4.3 mg)、**9**(550.0 mg)和**10**(3.0 mg)。

1.4 α -葡萄糖苷酶抑制活性测定方法

本测试在紫外分光光度计上进行,反应体系参照Jong-Anurakkun等^[10]的方法,优化后的测试方法为:0.5 mL的磷酸钾缓冲液(0.1 mmol L⁻¹, pH=6.8),加入100 μ L的 α -葡萄糖苷酶(0.2 U mL⁻¹)和0.5 mL样品溶液混匀。在37°C恒温15 min后,再加入0.5 mL的PNPG(2.5 mmol L⁻¹),混匀后37°C恒温15 min。最后加入1 mL的Na₂CO₃溶液(0.2 mol L⁻¹)终止反应。以阿卡波糖作为阳性对照,于405 nm波长下测反应液的OD值,重复3次取平均值。计算样品对 α -葡萄糖苷酶的抑制率:抑制率(%)=[A_{空白}-(A_{样品}-A_{背景})]/A_{空白}×100%。A_{空白}为不加待测样品反应后的吸光值,A_{样品}为加入待测样品反应后的吸光值,A_{背景}为只加待测样品反应后的吸光值。

1.5 结构鉴定

3-Hydroxy-1-(4-hydroxy-3,5-dimethoxyphenyl)propan-1-one (1) 白色粉末;ESI-MS m/z : 249.1 [M + Na]⁺, 225.3 [M - H]⁻;推断分子式为C₁₁H₁₄O₅; ¹H MNR (CDCl₃, 500 MHz): δ 7.26 (2H, d, J = 2.4 Hz, H-2, H-6), 4.04 (2H, t, J = 5.4 Hz, H-9), 3.96 (6H, s, 3, 5, 2×OCH₃), 3.20 (2H, t, J = 5.4 Hz, H-8); ¹³C NMR (CDCl₃, 125 MHz): δ 128.7 (C-1), 105.8 (C-2, 6), 147.2 (C-3, C-5), 140.5 (C-4), 199.3 (C-7), 40.3 (C-8), 58.7 (C-9), 56.9 (3, 5, 2×OCH₃)。以上波谱数据与文献[11]报道基本一致,故鉴定为3-hydroxy-1-(4-hydroxy-3,5-dimethoxyphenyl)propan-1-one。

6-Hydroxy-1,3,5,7-tetramethoxy-9H-xanthen-

9-one (2) 红色粉末; ESI-MS m/z : 333.2 $[M + H]^+$; 推断分子式为 $C_{17}H_{16}O_7$; 1H NMR ($CDCl_3$, 500 MHz): δ 7.50 (1H, s, H-8), 6.60 (1H, d, $J = 2.3$ Hz, H-4), 6.38 (1H, d, $J = 2.3$ Hz, H-2), 4.12 (3H, s, 5-OCH₃), 4.11 (3H, s, 1-OCH₃), 3.99 (3H, s, 3-OCH₃), 3.94 (3H, s, 7-OCH₃); ^{13}C NMR ($CDCl_3$, 125 MHz): δ 161.7 (C-1), 95.3 (C-2), 164.4 (C-3), 92.7 (C-4), 159.5 (C-4a), 134.0 (C-5), 143.8 (C-6), 144.5 (C-7), 100.7 (C-8), 115.6 (C-8a), 174.5 (C-9), 106.7 (C-9a), 144.5 (C-10a), 56.4 (1-OCH₃), 56.3 (3-OCH₃), 61.6 (5-OCH₃), 55.7 (7-OCH₃)。以上波谱数据与文献[12]报道基本一致, 故鉴定为 6-hydroxy-1,3,5,7-tetramethoxy-9H-xanthen-9-one。

2,6,2',6'-Tetramethoxy-4,4'-bis(2,3-epoxy-1-hydroxypropyl)biphenyl (3) 无色油状物; ESI-MS m/z : 441.3 $[M + Na]^+$; 推断分子式为 $C_{22}H_{26}O_8$; 1H NMR ($CDCl_3$, 500 MHz): δ 6.61 (4H, s, H-3, 3', 5, 5'), 5.52 (2H, s, 7, 7', 2 \times OH), 4.75 (2H, d, $J = 4.2$ Hz, H-7, 7'), 4.30 (2H, m, H-9H_b, H_b), 3.93 (12H, s, 2, 2', 6, 6', 4 \times OCH₃), 3.90 (2H, m, H-9H_a, H_a), 3.12 (2H, m, H-8, 8'); ^{13}C NMR ($CDCl_3$, 125 MHz): δ 131.9 (C-1, 1'), 147.0 (C-2, 2', 6, 6'), 102.5 (C-3, 3', 5, 5'), 134.1 (C-4, 4'), 86.0 (C-7, 7'), 54.2 (C-8, 8'), 71.7 (C-9, 9'), 56.3 (2, 2', 6, 6', 4 \times OCH₃)。以上波谱数据与文献[13]报道基本一致, 故鉴定为 2,6,2',6'-tetramethoxy-4,4'-bis-(2,3-epoxy-1-hydroxypropyl)biphenyl。

Cleomiscosin D (4) 白色粉末; ESI-MS m/z : 439.2 $[M + Na]^+$; 推断分子式为 $C_{21}H_{20}O_9$; 1H NMR ($CDCl_3$, 500 MHz): δ 7.95 (1H, d, $J = 9.5$ Hz, H-4), 6.90 (1H, s, H-5), 6.73 (2H, s, H-2', 6'), 6.33 (1H, d, $J = 9.5$ Hz, H-3), 4.95 (1H, d, $J = 8.0$ Hz, H-7'), 4.36 (1H, m, H-8'), 3.77 (6H, s, 3', 5', 2 \times OCH₃), 3.75 (3H, s, 4-OCH₃), 3.63 (1H, m, H-9a), 3.39 (1H, m, H-9b); ^{13}C NMR ($CDCl_3$, 125 MHz): δ 160.1 (C-2), 113.3 (C-3), 144.7 (C-4), 100.8 (C-5), 145.3 (C-6), 137.1 (C-7), 131.5 (C-8), 138.2 (C-9), 111.3 (C-10), 125.8 (C-1'), 105.7 (C-2', 6'), 148.0 (C-3', 5'), 136.3 (C-4'), 76.6 (C-7'), 77.8 (C-8'), 59.9 (C-9'), 55.9 (4-OCH₃), 56.2 (3', 5', 2 \times OCH₃)。以上波谱数据与文献[14]报道基本一致, 故鉴定为 cleomiscosin D。

Chuktabularin A (5) 白色无定形粉末; $[\alpha]_D^{28} + 238^\circ$ (c 0.1, $CHCl_3$), ESI-MS m/z : 799.4 $[M + Na]^+$; 推断分子式为 $C_{38}H_{48}O_{17}$; 1H NMR ($CDCl_3$, 500 MHz): δ 7.65 (1H, br s, H-21), 7.37 (1H, br t, $J =$

1.4 Hz, H-23), 6.50 (1H, br d, $J = 1.4$ Hz, H-22), 6.16 (1H, s, H-17), 5.67 (1H, d, $J = 3.3$ Hz, H-11), 5.63 (1H, d, $J = 3.3$ Hz, H-12), 5.23 (1H, s, H-3), 4.64 (1H, s, 1-OH), 4.63 (1H, s, H-30), 3.62 (3H, s, 7-OCH₃), 3.27 (1H, s, 9-OH), 3.10 (1H, dd, $J = 11.7, 7.6$ Hz, H-14), 2.58 (1H, br d, $J = 12.1$ Hz, H-5), 2.53 (1H, dd, $J = 11.7, 7.6$ Hz, H-15 β), 2.47 (3H, s, 3-OAc), 2.44 (1H, br d, $J = 16.5$ Hz, H-6a), 2.22 (1H, dd, $J = 16.5, 12.1$ Hz, H-6b), 2.10 (3H, s, 17-OAc), 2.08 (3H, s, 12-OAc), 2.07 (3H, s, 2-OAc), 1.94 (3H, s, 11-OAc), 1.90 (1H, d, $J = 11.7$ Hz, H-15 α), 1.84 (1H, d, $J = 11.3$ Hz, H-29a), 1.80 (1H, d, $J = 11.3$ Hz, H-29b), 1.63 (3H, s, H-32), 1.19 (3H, s, H-19), 0.91 (3H, s, H-18), 0.77 (3H, s, H-28); ^{13}C NMR ($CDCl_3$, 125 MHz): δ 84.6 (C-1), 82.3 (C-2), 83.5 (C-3), 45.5 (C-4), 41.3 (C-5), 34.3 (C-6), 173.3 (C-7), 90.2 (C-8), 76.3 (C-9), 52.7 (C-10), 72.5 (C-11), 73.4 (C-12), 41.8 (C-13), 44.9 (C-14), 35.6 (C-15), 71.3 (C-17), 19.1 (C-18), 18.9 (C-19), 122.5 (C-20), 140.7 (C-21), 109.7 (C-22), 143.2 (C-23), 16.2 (C-28), 40.1 (C-29), 71.2 (C-30), 110.4 (C-31), 17.8 (C-32), 51.9 (7-OCH₃), 2-OAc [(169.8), (21.1)], 3-OAc [(169.6), (21.2)], 11-OAc [(169.6), (21.1)], 12-OAc [(169.3), (20.7)], 17-OAc [(168.9), (20.6)]。以上波谱数据与文献[15]报道基本一致, 故鉴定为 chuktabularin A。

Chuktabularin B (6) 白色无定形粉末; $[\alpha]_D^{28} + 135^\circ$ (c 0.1, $CHCl_3$), ESI-MS m/z : 783.3 $[M + Na]^+$; 推断分子式为 $C_{37}H_{44}O_{17}$; 1H NMR ($CDCl_3$, 500 MHz): δ 7.53 (1H, br s, H-21), 7.45 (1H, br t, $J = 1.3$ Hz, H-23), 6.45 (1H, br d, $J = 1.3$ Hz, H-22), 6.09 (1H, s, H-17), 5.66 (1H, d, $J = 3.6$ Hz, H-11), 5.45 (1H, d, $J = 3.6$ Hz, H-12), 5.31 (1H, s, H-3), 4.96 (1H, d, $J = 12.5$ Hz, H-19a), 4.79 (1H, s, 1-OH), 4.61 (1H, s, H-30), 4.16 (1H, d, $J = 12.5$ Hz, H-19b), 3.37 (1H, s, 9-OH), 3.29 (1H, dd, $J = 11.7, 8.0$ Hz, H-14), 2.56 (1H, dd, $J = 11.7, 8.0$ Hz, H-15 β), 2.45 (3H, s, 3-OAc), 2.29 (1H, m, H-6), 2.11 (1H, d, $J = 11.7$ Hz, H-29a), 2.10 (3H, s, 11-OAc), 2.09 (3H, s, 2-OAc), 2.09 (3H, s, 12-OAc), 2.09 (3H, s, 17-OAc), 2.05 (1H, m, H-5), 2.01 (1H, dd, $J = 11.7, 11.7$ Hz, H-15 α), 1.99 (1H, d, $J = 11.7$ Hz, H-29b), 1.65 (3H, s, H-32), 0.91 (3H, s, H-18), 0.90 (3H, s, H-28); ^{13}C NMR ($CDCl_3$, 125 MHz): δ 85.2 (C-1), 80.9 (C-2), 82.6 (C-3), 45.2 (C-4), 40.5 (C-5), 30.9 (C-6), 173.1 (C-7), 89.6 (C-8),

75.2 (C-9), 52.1 (C-10), 71.4 (C-11), 72.0 (C-12), 41.1 (C-13), 44.2 (C-14), 35.7 (C-15), 71.3 (C-17), 19.4 (C-18), 68.7 (C-19), 122.0 (C-20), 140.0 (C-21), 109.0 (C-22), 142.9 (C-23), 15.0 (C-28), 38.4 (C-29), 71.1 (C-30), 110.9 (C-31), 18.7 (C-32), 2-OAc [(169.8), (20.8)], 3-OAc [(169.4), (21.1)], 11-OAc [(170.9), (20.4)], 12-OAc [(170.0), (20.6)], 17-OAc [(169.1), (20.3)]。以上波谱数据与文献[15]报道基本一致, 故鉴定为chuktabularin B。

Chubularisin H (7) 白色无定形粉末; $[\alpha]_D^{28} +116^\circ$ (*c* 0.1, CHCl₃), ESI-MS *m/z*: 911.5 [M + Na]⁺; 推断分子式为C₄₃H₅₂O₂₀; ¹H NMR (CDCl₃, 500 MHz): δ 7.48 (1H, br t, *J* = 1.7 Hz, H-21), 7.39 (1H, br t, *J* = 1.7 Hz, H-23), 7.06 (1H, br d, *J* = 3.0 Hz, H-15), 6.51 (1H, br d, *J* = 1.7 Hz, H-22), 6.43 (1H, s, H-17), 5.92 (1H, s, H-6), 5.46 (1H, s, H-3), 5.36 (1H, s, H-30), 5.32 (1H, br d, *J* = 3.5 Hz, H-12), 4.22 (1H, d, *J* = 3.5 Hz, H-11), 3.79 (3H, s, 7-OCH₃), 3.49 (1H, s, 2-OH), 2.91 (1H, m, H-2'), 2.88 (1H, s, H-5), 2.85 (1H, s, 1-OH), 2.65 (1H, dd, *J* = 6.8, 3.0 Hz, H-18a), 2.51 (1H, m, H-2''), 2.22 (3H, s, 6-OAc), 2.20 (3H, s, 3-OAc), 2.15 (1H, d, *J* = 10.9 Hz, H-29a), 1.95 (1H, d, *J* = 10.9 Hz, H-29b), 1.67 (3H, s, 12-OAc), 1.66 (3H, s, H-32), 1.43 (1H, d, *J* = 6.8 Hz, H-18b), 1.34 (3H, s, H-19), 1.31 (3H, d, *J* = 7.3 Hz, H-4'), 1.25 (3H, d, *J* = 6.6 Hz, H-3'), 1.19 (3H, d, *J* = 6.9 Hz, H-4''), 1.17 (3H, d, *J* = 6.9 Hz, H-3''), 1.00 (3H, s, H-28); ¹³C NMR (CDCl₃, 125 MHz): δ 83.1 (C-1), 76.7 (C-2), 86.0 (C-3), 44.9 (C-4), 43.1 (C-5), 70.8 (C-6), 171.7 (C-7), 78.3 (C-8), 90.8 (C-9), 45.1 (C-10), 74.9 (C-11), 66.4 (C-12), 31.1 (C-13), 30.9 (C-14), 69.3 (C-15), 167.0 (C-16), 71.4 (C-17), 18.6 (C-18), 15.2 (C-19), 122.3 (C-20), 142.1 (C-21), 109.8 (C-22), 143.5 (C-23), 15.4 (C-28), 40.1 (C-29), 70.0 (C-30), 119.6 (C-31), 16.3 (C-32), 53.7 (7-OCH₃), 15-isobutyryloxyl [177.9 (C-1'), 34.2 (C-2'), 19.8 (C-3'), 17.9 (C-4')], 30-isobutyryloxyl [173.4 (C-1''), 33.9 (C-2''), 19.5 (C-3''), 18.9 (C-4'')], 3-OAc [(169.2), (21.1)], 6-OAc [(169.2), (21.2)], 12-OAc [(170.7), (19.7)]。以上波谱数据与文献[16]报道基本一致, 故鉴定为chubularisin H。

Chubularisin I (8) 白色无定形粉末; $[\alpha]_D^{28} +124^\circ$ (*c* 0.1, CHCl₃), ESI-MS *m/z*: 839.3 [M + Na]⁺; 推断分子式为C₄₀H₄₈O₁₈; ¹H NMR (CDCl₃, 500 MHz): δ 7.47 (1H, br s, H-21), 7.39 (1H, br s, H-23), 7.21

(1H, br d, *J* = 2.8 Hz, H-15), 6.49 (1H, br s, H-22), 6.43 (1H, s, H-17), 5.52 (1H, s, H-3), 5.40 (1H, s, H-30), 5.13 (1H, br d, *J* = 3.2 Hz, H-12), 4.17 (1H, d, *J* = 3.2 Hz, H-11), 3.80 (1H, s, 2-OH), 3.76 (3H, s, 7-OCH₃), 2.92 (1H, m, H-2'), 2.84 (1H, s, 1-OH), 2.65 (1H, d, *J* = 16.7 Hz, H-6a), 2.64 (1H, dd, *J* = 6.8, 2.8 Hz, H-18a), 2.58 (1H, d, *J* = 12.4 Hz, H-5), 2.45 (1H, d, *J* = 16.7 Hz, H-6b), 2.32 (2H, q, *J* = 7.7 Hz, H-2''), 2.20 (3H, s, 3-OAc), 1.91 (2H, s, H-29), 1.66 (3H, s, H-32), 1.66 (3H, s, 12-OAc), 1.43 (1H, d, *J* = 6.8 Hz, H-18b), 1.37 (3H, d, *J* = 7.0 Hz, H-4'), 1.30 (3H, s, H-19), 1.24 (3H, d, *J* = 7.0 Hz, H-3'), 1.19 (3H, t, *J* = 7.1 Hz, H-3''), 0.82 (3H, s, H-28); ¹³C NMR (CDCl₃, 125 MHz): δ 83.1 (C-1), 76.5 (C-2), 85.8 (C-3), 45.0 (C-4), 38.2 (C-5), 33.1 (C-6), 173.9 (C-7), 78.5 (C-8), 90.6 (C-9), 44.1 (C-10), 75.0 (C-11), 66.6 (C-12), 31.2 (C-13), 30.8 (C-14), 69.9 (C-15), 167.1 (C-16), 71.5 (C-17), 18.8 (C-18), 14.7 (C-19), 122.1 (C-20), 142.2 (C-21), 109.8 (C-22), 143.5 (C-23), 14.3 (C-28), 38.9 (C-29), 70.7 (C-30), 119.9 (C-31), 16.4 (C-32), 52.6 (7-OCH₃), 15-isobutyryl-oxyl [178.0 (C-1'), 34.2 (C-2'), 19.5 (C-3'), 17.8 (C-4')], 30-propionyloxyl [171.1 (C-1''), 27.4 (C-2''), 9.2 (C-3'')], 3-OAc [(169.3), (21.1)], 12-OAc [(170.8), (20.0)]。以上波谱数据与文献[16]报道基本一致, 故鉴定为chubularisin I。

Tabularisin A (9) 白色无定形粉末; $[\alpha]_D^{28} +185^\circ$ (*c* 0.1, CHCl₃), ESI-MS *m/z*: 883.4 [M + Na]⁺; 推断分子式为C₄₁H₄₈O₂₀; ¹H NMR (CDCl₃, 500 MHz): δ 7.48 (1H, br s, H-21), 7.39 (1H, br s, H-23), 7.09 (1H, d, *J* = 3.0 Hz, H-15), 6.50 (1H, d, *J* = 2.0 Hz, H-22), 6.43 (1H, s, H-17), 5.90 (1H, s, H-6), 5.46 (1H, s, H-3), 5.36 (1H, s, H-30), 5.31 (1H, br d, *J* = 3.6 Hz, H-12), 4.22 (1H, d, *J* = 3.6 Hz, H-11), 3.79 (3H, s, 7-OCH₃), 3.36 (1H, s, 2-OH), 2.87 (1H, s, H-5), 2.83 (1H, s, 1-OH), 2.66 (1H, dd, *J* = 6.8, 3.0 Hz, H-18a), 2.50~2.55 (1H, m, H-2'), 2.33 (3H, s, 15-OAc), 2.21 (3H, s, 6-OAc), 2.18 (3H, s, 3-OAc), 2.15 (1H, d, *J* = 10.9 Hz, H-29b), 1.94 (1H, d, *J* = 10.9 Hz, H-29a), 1.66 (3H, s, H-32), 1.66 (3H, s, 12-OAc), 1.42 (1H, d, *J* = 6.8 Hz, H-18b), 1.33 (3H, s, H-19), 1.19 (3H, d, *J* = 7.2 Hz, H-4'), 1.18 (3H, d, *J* = 7.2 Hz, H-3'), 0.98 (3H, s, H-28); ¹³C NMR (CDCl₃, 125 MHz): δ 83.0 (C-1), 76.6 (C-2), 85.9 (C-3), 44.8 (C-4), 43.1 (C-5), 70.7 (C-6), 171.6 (C-7), 78.2 (C-8), 90.7 (C-9), 45.1 (C-10),

74.9 (C-11), 66.3 (C-12), 31.0 (C-13), 30.7 (C-14), 69.7 (C-15), 166.9 (C-16), 71.4 (C-17), 18.6 (C-18), 15.2 (C-19), 122.1 (C-20), 142.1 (C-21), 109.7 (C-22), 143.5 (C-23), 15.3 (C-28), 40.0 (C-29), 70.1 (C-30), 119.6 (C-31), 16.3 (C-32), 53.7 (7-OCH₃), 30-isobutyryloxy [173.5 (C-1'), 34.0 (C-2'), 19.5 (C-3'), 18.9 (C-4')], 3-OAc [(169.1), (21.1)], 6-OAc [(169.2), (21.1)], 12-OAc [(170.7), (19.7)], 15-OAc [(172.3), (21.6)]。以上波谱数据与文献[17]报道基本一致, 故鉴定为tabularisin A。

Tabularisin B (10) 白色无定形粉末; $[\alpha]_D^{28} +264^\circ$ (c 0.1, CHCl₃), ESI-MS m/z : 841.4 [M + Na]⁺; 推断分子式为C₃₉H₄₆O₁₉; ¹H NMR (CDCl₃, 500 MHz): δ 7.69 (1H, br s, H-21), 7.53 (1H, br s, H-23), 7.08 (1H, d, $J = 2.8$ Hz, H-15), 6.56 (1H, br s, H-22), 6.42 (1H, s, H-17), 5.47 (1H, s, H-3), 5.36 (1H, s, H-30), 5.34 (1H, s, H-6), 4.25 (1H, d, $J = 3.5$ Hz, H-11), 4.03 (1H, d, $J = 3.5$ Hz, H-12), 3.81 (3H, s, 7-OCH₃), 3.38 (1H, s, 2-OH), 2.93 (1H, s, 1-OH), 2.77 (1H, s, H-5),

2.50~2.55 (1H, m, H-2'), 2.46 (1H, dd, $J = 6.7, 2.8$ Hz, H-18a), 2.33 (3H, s, 15-OAc), 2.23 (3H, s, 6-OAc), 2.18 (3H, s, 3-OAc), 2.14 (1H, d, $J = 10.8$ Hz, H-29b), 1.96 (1H, d, $J = 10.8$ Hz, H-29a), 1.67 (3H, s, H-32), 1.37 (1H, m, H-18b), 1.34 (3H, s, H-19), 1.21 (3H, d, $J = 7.0$ Hz, H-4'), 1.19 (3H, d, $J = 7.0$ Hz, H-3'), 0.97 (3H, s, H-28); ¹³C NMR (CDCl₃, 125 MHz): δ 83.2 (C-1), 76.4 (C-2), 85.8 (C-3), 44.8 (C-4), 42.9 (C-5), 71.1 (C-6), 171.4 (C-7), 77.9 (C-8), 91.1 (C-9), 45.3 (C-10), 76.5 (C-11), 65.1 (C-12), 34.7 (C-13), 31.7 (C-14), 69.8 (C-15), 167.1 (C-16), 71.4 (C-17), 18.3 (C-18), 15.2 (C-19), 122.1 (C-20), 142.7 (C-21), 108.9 (C-22), 145.0 (C-23), 15.4 (C-28), 39.9 (C-29), 70.0 (C-30), 119.4 (C-31), 16.3 (C-32), 53.9 (7-OCH₃), 30-isobutyryloxy [173.6 (C-1'), 34.0 (C-2'), 19.6 (C-3'), 18.9 (C-4')], 3-OAc [(169.1), (21.2)], 6-OAc [(169.6), (21.2)], 15-OAc [(172.4), (21.6)]。以上波谱数据与文献[17]报道基本一致, 故鉴定为tabularisin B。

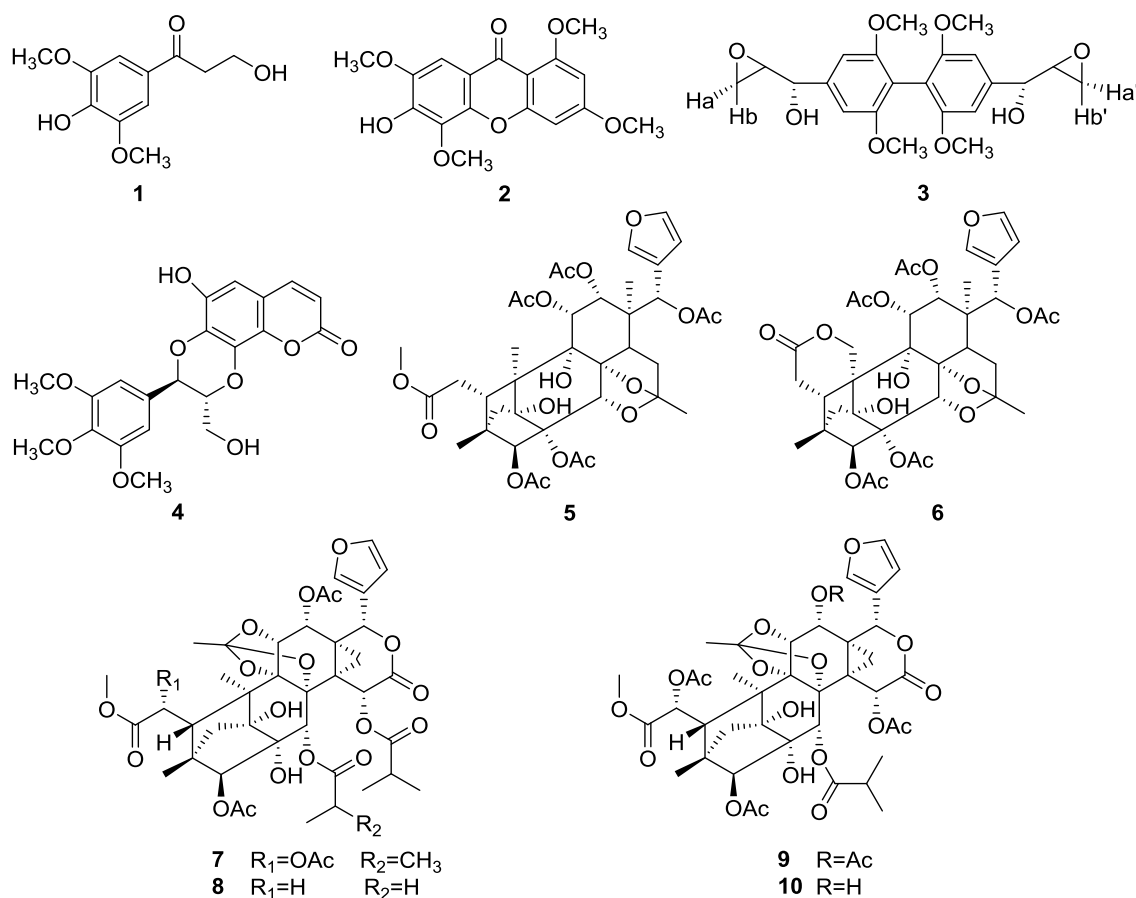


图1 化合物1~10的结构

Fig. 1 Structures of compounds 1-10

1.6 化合物对 α -葡萄糖苷酶抑制活性测试

本试验采用体外抑制法测定了化合物**1~10**的 α -葡萄糖苷酶抑制活性,结果表明,麻楝中柠檬苦素类化合物表现出明显的对 α -葡萄糖苷酶

的抑制活性,且部分柠檬苦素类化合物的活性优于阳性对照阿卡波糖;另外,部分木脂素类化合物也表现出一定的 α -葡萄糖苷酶的抑制活性(表1)。

表1 化合物的 α -葡萄糖苷酶抑制活性(IC₅₀)

Table 1 α -Glucosidase inhibitory activity of compounds (IC₅₀)

化合物 Compound	IC ₅₀ ($\mu\text{g mL}^{-1}$)	化合物 Compound	IC ₅₀ ($\mu\text{g mL}^{-1}$)
1	353.2	7	505.1
2	362.2	8	—
3	—	9	377.5
4	—	10	—
5	—	阿卡波糖 Acarbose	794.5
6	338.0		

2 结果和讨论

本文采用多种色谱技术,从麻楝枝干提取物中分离得到了10个化合物,分别鉴定为: 3-hydroxy-1-(4-hydroxy-3,5-dimethoxyphenyl)propan-1-one (**1**)、6-hydroxy-1,3,5,7-tetramethoxy-9H-xanthen-9-one (**2**)、2,6,2',6'-tetramethoxy-4,4'-bis(2,3-epoxy-1-hydroxypropyl)biphenyl (**3**)、cleomiscosin D (**4**)、chuktabularin A (**5**)、chuktabularin B (**6**)、chubularisin H (**7**)、chubularisin I (**8**)、tabularisin A (**9**)和tabularisin B (**10**),其中有6个为柠檬苦素类化合物,化合物**1~4**为首次从麻楝属植物中分离得到。对所得化合物进行了体外 α -葡萄糖苷酶抑制活性测试,结果表明,化合物**1**、**2**、**6**、**7**和**9**均具有较好的抑制活性,本文为首次报道柠檬苦素类化合物具有体外抑制 α -葡萄糖苷酶的活性。同时,据文献报道,化合物**1**具有抗氧化活性^[18]和细胞毒活性^[19],化合物**4**具有抗炎活性^[20],化合物**7**具有较强的钾离子通道抑制活性^[16],但并未见其他化合物生物活性的报道。本研究结果为进一步挖掘麻楝的药用价值提供了科学依据。

参考文献

- CHEN S K, CHEN B Y, LI H. Flora Reipublicae Popularis Sinicae, Tomus 43(3) [M]. Beijing: Science Press, 1997: 47–49.
陈书坤, 陈邦余, 李恒. 中国植物志, 第43卷第3分册 [M]. 北京: 科学出版社, 1997: 47–69.
- TAN Q G, LUO X D. Meliaceae limonoids: Chemistry and biological activities [J]. Chem Rev, 2011, 111(11): 7437–7522. doi: 10.1021/cr9004023.
- LUO J, WANG J S, LUO J G, et al. Velutabularins A–J, phragmalin-type limonoids with novel cyclic moiety from *Chukrasia tabularis* var. *velutina* [J]. Tetrahedron, 2011, 67(16): 2942–2948. doi: 10.1016/j.tet.2011.02.049.
- LUO J, LI Y, WANG J S, et al. D-ring-opened phragmalin-type limonoids from *Chukrasia tabularis* var. *velutina* [J]. Chem Biod, 2011, 8(12): 2261–2269. doi: 10.1002/cbdv.201000285.
- LIU H B, ZHANG H, LI P, et al. Chukrasones A and B: Potential Kv1.2 potassium channel blockers with new skeletons from *Chukrasia tabularis* [J]. Org Lett, 2012, 14(17): 4438–4441. doi: 10.1021/ol301942v.
- ZHANG C R, YANG S P, CHEN X Q, et al. Limonoids from the twigs and leaves of *Chukrasia tabularis* [J]. Hel Chim Acta, 2008, 91(12): 2338–2350. doi: 10.1002/hlca.200890254.
- YIN J L, DI Y T, FANG X, et al. Tabulvelutin A, the first 19-nor limonoid with unprecedented ring system from *Chukrasia tabularis* var. *velutina* [J]. Tetrah Lett, 2011, 52(24): 3083–3085. doi: 10.1016/j.tetlet.2011.03.112.
- NAKATANI M, ABDELGALEIL S A M, SAAD M M G, et al. Phragmalin limonoids from *Chukrasia tabularis* [J]. Phytochemistry, 2004, 65(20): 2833–2841. doi: 10.1016/j.phytochem.2004.08.010.
- LUO J, WANG J S, WANG X B, et al. Phragmalin-type limonoid orthoesters from *Chukrasia tabularis* var. *velutina* [J]. Chem Pharm Bull, 2011, 59(2): 225–230. doi: 10.1248/cpb.59.225.
- JONG-ANURAKKUN N, BHANDARI M R, KAWABATA J. α -Glucosidase inhibitors from devil tree (*Alstonia scholaris*) [J]. Food Chem, 2007, 103(4): 1319–1323. doi: 10.1016/j.foodchem.2006.10.043.
- MA Z J, ZHAO Z J. Studies on chemical constituents from stem barks of *Fraxinus paxiana* [J]. China J Chin Mat Med, 2008, 33(16):

- 1990–1993. doi: 10.3321/j.issn:1001-5302.2008.16.016.
- 马志静, 赵志娟. 秦岭白蜡树化学成分的研究 [J]. 中国中药杂志, 2008, 33(16): 1990–1993. doi: 10.3321/j.issn:1001-5302.2008.16.016.
- [12] KIJJOA A, GONZALEZ M J, AFONSO C M, et al. Xanthones from *Calophyllum teysmannii* var. *inophylloide* [J]. Phytochemistry, 2000, 53(8): 1021–1024. doi: 10.1016/S0031-9422(99)00520-8.
- [13] DAY S H, WANG J P, WON S J, et al. Bioactive constituents of the roots of *Cynanchum atratum* [J]. J Nat Prod, 2001, 64(5): 608–611. doi: 10.1021/np000428b.
- [14] KUMAR S, RAY A B, KONNO C, et al. Cleomiscosin D, a coumarinolignan from seeds of *Cleome viscosa* [J]. Phytochemistry, 1988, 27(2): 636–638. doi: 10.1016/0031-9422(88)83163-7.
- [15] ZHANG C R, YANG S P, LIAO S G, et al. Chuktabularins A-D, four new limonoids with unprecedented carbon skeletons from the stem bark of *Chukrasia tabularis* [J]. Org Lett, 2007, 9(17): 3383–3386. doi: 10.1021/ol701437h.
- [16] LIU H B, ZHANG H, LI P, et al. Kv1.2 potassium channel inhibitors from *Chukrasia tabularis* [J]. Org Biomol Chem, 2012, 10(7): 1448–1458. doi: 10.1039/C1OB06666H.
- [17] FAN C Q, WANG X N, YIN S, et al. Tabularisins A–D, phragmalin ortho esters with new skeleton isolated from the seeds of *Chukrasia tabularis* [J]. Tetrahedron, 2007, 63(29): 6741–6747. doi: 10.1016/j.tet.2007.04.078.
- [18] ASIKIN Y, TAKAHASHI M, MISHIMA T, et al. Antioxidant activity of sugarcane molasses against 2,2'-azobis (2-aminopropane) dihydrochloride-induced peroxy radicals [J]. Food Chem, 2013, 141(1): 466–472. doi: 10.1016/j.foodchem.2013.03.045.
- [19] KIM K H, MOON E, CHOI S U, et al. Biological evaluation of phenolic constituents from the trunk of *Berberis koreana* [J]. Bioorg Med Chem Lett, 2011, 21(8): 2270–2273. doi: 10.1016/j.bmcl.2011.02.104.
- [20] CHEN J J, WANG T Y, HWANG T L. Neolignans, a coumarinolignan, lignan derivatives, and a chromene: Anti-inflammatory constituents from *Zanthoxylum avicennae* [J]. J Nat Prod, 2008, 71(2): 212–217. doi: 10.1021/np070594k.