

枇杷果肉有机酸组分及有机酸在果实内的分布

陈发兴^{a,b}, 刘星辉^a, 陈立松^{a,c*}

(福建农林大学, a. 园艺学院; b. 亚热带果树研究所; c. 园艺植物生理生化与分子生物学研究所, 福州 350002)

摘要: 用高效离子交换色谱(HPIC)测定了枇杷(*Eriobotrya japonica* Lindl.)18个品种(小毛枇杷、夹脚、卓南1号、解放钟、富阳、森尾早生、华宝2号、香钟10号、白花、土肥、多宝2号、乌躬白、洛阳青、茂木、早钟6号、白梨、塘头4号和长红3号)的成熟果肉和2个品种(解放钟和早钟6号)成熟果实不同组织有机酸含量。结果表明,成熟果肉中均含有苹果酸、奎尼酸、柠檬酸、异柠檬酸、 α -酮戊二酸、富马酸、草酰乙酸、酒石酸8种有机酸,有的还含有微量的阿魏酸、顺乌头酸和 β -香豆酸。大多数品种果肉中苹果酸含量最高,平均含量为4 399 mg kg⁻¹ FW,占总酸的62.7%;其次是奎尼酸,其平均含量为2 042 mg kg⁻¹ FW,占总酸的29.1%。品种之间可滴定酸和有机酸含量差异很大。通过对果肉可滴定酸进行聚类分析,可把18个枇杷品种分为五个组群:极高酸(小毛枇杷)、高酸(夹脚、卓南1号、解放钟和富阳)、中酸(森尾早生、华宝2号、香钟10号、白花、土肥和多宝2号)、低酸(乌躬白、洛阳青、茂木和早钟6号)和极低酸(白梨、塘头4号和长红3号)。解放钟和早钟6号果肉和果皮的总酸含量及可滴定酸均无显著差异,但果皮和果肉的总酸含量和可滴定酸均大大高于种子。相似于果肉,果皮和种子的主要有机酸也是苹果酸和奎尼酸。果皮中苹果酸含量远高于奎尼酸,但种子中苹果酸含量比奎尼酸稍低。此外,种子中苹果酸和奎尼酸比果肉和果皮中的低得多。

关键词: 枇杷; 果实; 苹果酸; 有机酸组分; 果肉; 奎尼酸; 种子; 果皮

中图分类号: Q946.81

文献标识码: A

文章编号: 1005-3395(2008)03-0236-08

Organic Acid Composition in the Pulp of Loquat (*Eriobotrya japonica* Lindl.) and Distribution in Fruits

CHEN Fa-xing^{a,b}, LIU Xing-hui^a, CHEN Li-song^{a,c*}

(a. College of Horticulture; b. Institute of Subtropical Fruits; c. Institute of Horticultural Plant Physiology, Biochemistry and Molecular Biology, Fujian Agriculture and Forestry University, Fuzhou 350002, China)

Abstract: Organic acids from the ripe pulp of 18 loquat (*Eriobotrya japonica* Lindl.) cultivars ('Xiaomaopipa', 'Jiajiao', 'Zhuonan 1', 'Jiefangzhong', 'Fuyang', 'Moriowase', 'Huabao 2', 'Xiangzhong 10', 'Baihua', 'Toi', 'Duobao 2', 'Wugongbai', 'Luoyangchin', 'Mogi', 'Zaozhong 6', 'Baili', 'Tantou 4', 'Changhong 3') and their distribution in the ripe fruit of 2 cultivars ('Jiefangzhong' and 'Zaozhong 6') were determined by high-performance ion-exchange chromatography (HPIC). Eight organic acids (malic, quinic, citric, iso-citric, α -ketoglutaric, fumaric, oxaloacetic and tartaric acids) were identified in ripe pulp, while trace quantities of ferulic, *cis*-aconitic and β -coumaric acids were identified in several cultivars. The predominant organic acid of ripe pulp of most cultivars was malic acid, with an average content of 4 399 mg kg⁻¹ FW for all cultivars, accounting for 62.7% of the total acids. Quinic acid was the second in abundance, with an average content of 2 042 mg kg⁻¹ FW, accounting for 29.1% of the total acids. Other acids ranged from trace to 3.0% of the total acids. Considerable

Received: 2007-07-05

Accepted: 2007-09-24

Foundation items: The National Natural Science Foundation of China (No. 30571292); Fujian Provincial Natural Science Foundation of Fujian Province, China (No. B0310008); Scientific Foundation from Fujian Provincial Department of Education, China (No. JA05231)

* Corresponding author

variations in titratable acidity and organic acid content exist among cultivars. Based on titratable acidity, 18 cultivars could be separated into 5 groups by clustering analysis: very high acidity ('Xiaomaopipa'), high acidity ('Jiajiao', 'Zhuonan 1', 'Jiefangzhong' and 'Fuyang'), medium acidity ('Moriowase', 'Huabao 2', 'Xiangzhong 10', 'Baihua', 'Toi', 'Duobao 2'), low acidity ('Wugongbai', 'Luoyangchin', 'Mogi' and 'Zaozhong 6'), and very low acidity ('Baili', 'Tantou 4' and 'Changhong 3'). No significant difference was found in the absolute amounts of total acids and titratable acidity between pulp and skins, while the amounts of total acids and titratable acidity in seeds was far lower than those in pulp and skins. The major organic acids identified in skins and seeds were malic and quinic acids. In skins, the content of malic acid was far higher than quinic acid, but in seeds it was slightly lower than quinic acid. In addition, malic acid and quinic acid content in seeds was far lower than those in pulp and skins.

Key words: Loquat; Fruit; Malic acid; Organic acid composition; Pulp; Quinic acid; Seed; Skin

Many kinds of fruits accumulate considerable amounts of organic acids during various development stages. These compounds are used extensively as additives, namely antioxidants (tartaric, malic and citric acids) or preservatives (sorbic and benzoic acids)^[1-2]. Knowledge of the qualitative and quantitative distribution of organic acids in fruit is of considerable importance, since they influence fruit organoleptic and nutritional property. Organic acids also affect the nature and contents of other organic compounds in fruit.

Loquat (*Eriobotrya japonica* Lindl.) is a subtropical evergreen fruit tree and now commercially grown in many countries, including China, Japan, Italy, Brazil, Spain etc. In China, loquat is cultivated in 19 provinces, ranging from the Yangtze River to Hainan Island. There are many cultivars or selections in various provinces of China. The largest collection of germplasm, more than 250 cultivars, is located in Fuzhou, China^[3].

Information regarding the nature and amounts of organic acids of loquat fruit has been known^[4-8], but limited data are available on the organic acids of different loquat cultivars. Shaw and Wilson^[8] determined the organic acid composition in the fruit of 3 loquat cultivars ('Tanaka', 'Thales' and 'Christmas'). Although Rajput and Singh^[7] investigated the total acids of edible portion from 9 loquat cultivars, the quantities of individual acid present had not been determined. It has been known that organic acids are not evenly distributed within fruit^[9-11]. To our best knowledge, such data have not been reported for

loquat fruit. The objective of this study was to investigate organic acid profile of pulp from the ripe fruit of 18 loquat cultivars grown in the Germplasm Repository for Loquat in Fujian Fruit Research Institute, Fujian Academy of Agricultural Sciences located at Fuzhou, China. We also reported the distribution of organic acids within the ripe fruit of 2 loquat cultivars.

1 Materials and methods

1.1 Fruit materials

Eighteen loquat (*Eriobotrya japonica* Lindl.) cultivars were investigated in this study in 2004. All the samples were collected from 15-year old trees grown in the Germplasm Repository for Loquat in Fujian Fruit Research Institute, Fujian Academy of Agricultural Sciences located at Fuzhou, China. The trees were grown at a spacing of 3 m × 4 m. They received standard horticultural practices, and disease and insect control. Thirty ripe fruit from each cultivar were collected. The excised fruit were kept on ice and brought to laboratory. Fruit were carefully dissected into pulp, skins and seeds, frozen in liquid N₂ and then stored at -80°C until analysis.

1.2 Chemicals

All chemicals used were HPLC grade or analytical reagents. Quinic, *cis*-aconitic and oxaloacetic acid were purchased from Fluka Chemical Co.; L-Malic acid was purchased from Amresco Inc.; Tartaric, cinnamic and α -ketoglutaric acids were purchased from Shanghai Chemical Reagent Co. Ltd.; Fumaric, citric, β -coumaric, ferulic and iso-citric acids were

purchased from Sigma Chemical Co.

1.3 Analysis of titratable acidity

Titratable acidity was titrated with 0.1 mmol/L NaOH to the end point pH 8.1 and the total acidity calculated as malic acid^[12].

1.4 Determination of organic acids

Organic acids were extracted according to the method of Ding et al.^[5] with some modifications. One gram of frozen sample was ground in 3 ml of 95% (V/V) ice-cold methanol, shaken for 10 min, and filtered. The pellet was extracted twice again with 3 ml of 80% (V/V) ice-cold methanol. The combined extracts were evaporated under vacuum at less than

35°C until the methanol was removed, and the residue was dissolved in 25 ml water and centrifuged to remove undissolved materials. One ml of aliquot was passed through the 0.45 μm membrane filter, and the filtrate was then analysed by high-performance ion-exchange chromatography (HPIC, Dionex DX-320, Dionex Corp., Sunnyvale, California) according to the method of Chen et al.^[4]. Figs. 1 and 2 show the chromatograms of organic acids in loquat pulp and a standard mixture of organic acids, respectively.

2 Results and discussion

As shown in Table 1, ripe pulp from 18 cultivars all composed of, at least, 8 identified organic acids:

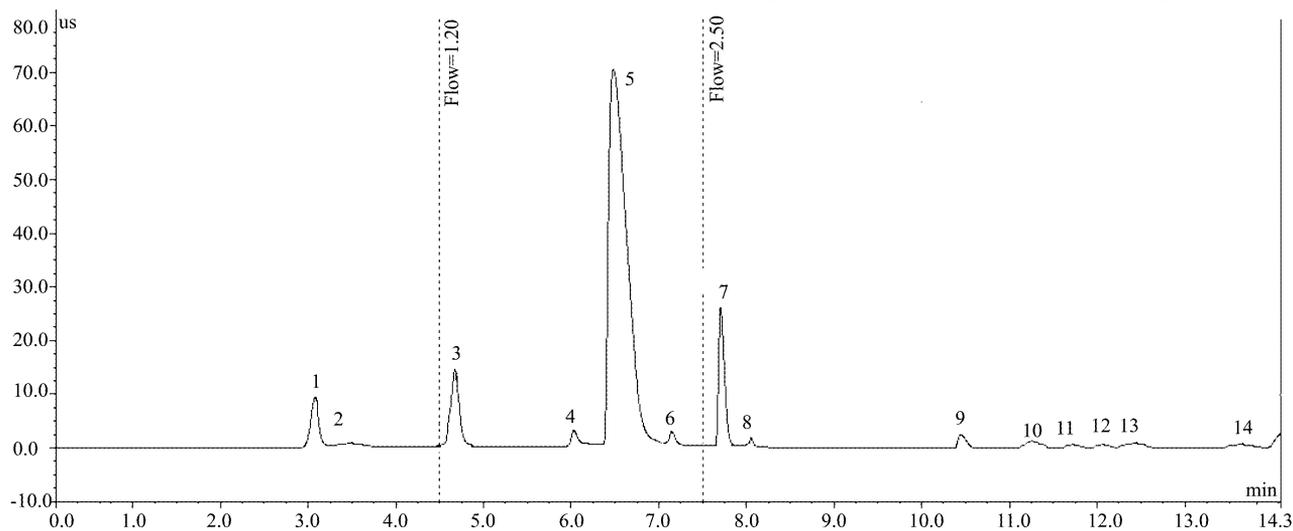


Fig. 1 Chromatogram of organic acids in pulp from 'Jiegfangzhong' loquat

1. Quinic acid; 2. Oxaloacetic acid; 3 ~ 4. Unknowns; 5. L-Malic acid; 6. Tartaric acid; 7. α -Ketoglutaric acid; 8. Fumaric acid; 9. Unknown; 10. Citric acid; 11. Iso-citric acid; 12. *cis*-Aconitic acid; 13. Ferulic acid; 14. β -Coumaric acid.

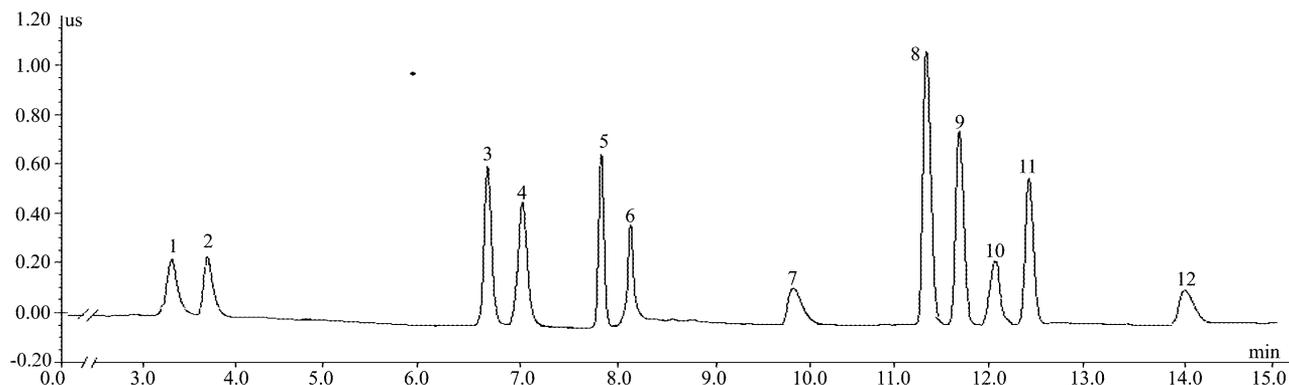


Fig. 2 Chromatogram of a standard mixture of organic acids

1. Quinic acid; 2. Oxaloacetic acid; 3. L-Malic acid; 4. Tartaric acid; 5. α -Ketoglutaric acid; 6. Fumaric acid; 7. Cinnamic acid; 8. Citric acid; 9. Iso-citric acid; 10. *cis*-Aconitic acid; 11. Ferulic acid; 12. β -Coumaric acid.

malic, quinic, citric, iso-citric, α -ketoglutaric, fumaric, oxaloacetic and tartaric acids. Ripe pulp of several cultivars also contained trace quantities of ferulic, *cis*-aconitic and β -coumaric acids. This is in general agreement with previous reports^[5-6,8,13]. As with other studies, we found that the predominant organic acid of pulp from the ripe fruit of major loquat cultivars was malic acid with an average content of 4 399 mg kg⁻¹ FW for all cultivars, accounting for 62.7% of the total acids. The highest malic acid content, 8 912 mg kg⁻¹ FW, was found in 'Xiaomaopipa' and the lowest content, 1 299 mg kg⁻¹ FW in 'Changhong 3'. Malic acid was reported to be 83% of the total acids and citric acid was the only other acid present in loquat fruit^[6]. Shaw and Wilson^[8] determined the organic acid composition in the fruit of 3 loquat cultivars ('Tanaka', 'Thales' and 'Christmas'). Malic acid was the predominant organic acid and citric acid was the second in the abundance. Two acids were quantified in all samples. In addition, trace quantities of several unknown acids was observed in 'Tanake' fruit. Randhawa and Singh^[13] reported the presence of succinic and tartaric acids in addition to malic and citric acids in loquat fruit, but no succinic acid was detected in our study (Table 1).

Our data showed clearly that quinic acid was the second abundant acid in ripe pulp. Quinic acid content ranged from 1 001 mg kg⁻¹ FW for 'Zaozhong 6' to 3 296 mg kg⁻¹ FW for 'Zhuonan 1', with an average content of 2 042 mg kg⁻¹ FW, accounting for 29.1% of the total acids. Except for 'Changhong 3' and 'Wugongbai', content of malic acid was much higher than that of quinic acid (Table 1). This is consistent with our previous report that loquat fruit and leaves contained large quantities of quinic acid^[4]. To our best knowledge, quinic acid had not been reported previously in loquat fruit by other workers. Quinic acid is formed by quinate dehydrogenase (EC 1.1.1.25) from 3-dehydroquinate^[14] and has been reported to present in large quantities in sea buckthorn (*Hippophae rhamnoides* L.)^[15], peach (*Prunus spersica*

(L.) Batsch)^[16-18], *Prunus davidiana* (Carr.) Franch.^[19], apple (*Malus domestica* Borkh.), pear (*Pyrus communis* L.)^[20-21], highbush blueberry (*Vaccinium corymbosum* L.), rabbiteye blueberry (*V. ashei* Reade)^[22], *V. acrostaphylos* L., *V. myrtilus* L.^[23], satsuma mandarin (*Citrus unshiu* Marc.)^[24], kiwifruit (*Actinidia chinensis* Planch.)^[10,25-27], and quince (*Cydonia oblonga* Miller)^[28]. Other acids in ripe pulp ranged from trace to 3.0% of the total acids (Table 1). The quantities of malic acid and total acids, and titratable acidity of pulp are similar to previous reports^[6-8].

As shown in Table 1, titratable acidity and contents of total acids and individual organic acid in ripe pulp depended largely upon cultivars. The highest coefficient of variation (CV), 158.8%, was observed in tartaric acid and the lowest CV, 27.9% in quinic acid. The CVs for titratable acidity, total acids and malic acid were 40.6%, 29.7% and 39.6%, respectively. Despite of this variation, there are features common to these analyzed samples. Except for 'Wugongbai' (78.2%) and 'Xiaomaopipa' (88.0%), the sum of malic acid and quinic acid represented 90.2% ~ 95.6% of the total acids in the other 16 cultivars, with an average of 91.9% and a CV of 4.3% (data not shown). Similar results have been obtained in quince (*Cydonia oblonga* Miller) pulp and peel^[28].

Table 2 shows the correlation coefficients among organic acids of ripe pulp. The highest relationship was observed between total acids and titratable acidity, with a correlation coefficient of 0.9644. The titratable acidity, total acids, malic acid and iso-citric acid were positively correlated with each other at $P < 0.001$. Citric acid was also positively correlated with titratable acidity ($P < 0.01$), total acids ($P < 0.01$), malic acid ($P < 0.05$), and iso-citric acid ($P < 0.001$). Quinic acid was correlated with titratable acidity, citric and iso-citric acids at $P < 0.05$, but it was not significantly correlated with total acids and malic acid. Oxaloacetic acid was positively correlated with titratable acidity ($P < 0.001$), total acids ($P < 0.01$), malic acid ($P < 0.01$), and iso-citric acid ($P < 0.01$).

Table 1 Organic acid composition (mg kg⁻¹ FW) in ripe pulp of 18 loquat cultivars

Cultivars	Malic	Quinic	Citric	Iso-citric	α -Ketoglutaric	Fumaric	Oxaloacetic	Tartaric	Ferulic <i>cis</i> -Aconitic	β -Coumaric	Total acids	Titratable acidity (%)
Xiaomaopipa	8912 \pm 92a	2490 \pm 85bc	114 \pm 2ab	38 \pm 3a	260 \pm 22a	101 \pm 14ab	126 \pm 17a	900 \pm 181a	7 \pm 0a	10 \pm 0a	12958 \pm 129a	1.270 \pm 0.119a
Jiajiao	5987 \pm 422c	2431 \pm 29bc	52 \pm 5c	17 \pm 1cd	259 \pm 17a	65 \pm 13bcd	81 \pm 10b	Trace	3 \pm 0b	7 \pm 0c	8902 \pm 416b	0.818 \pm 0.073b
Zhuanan 1	5433 \pm 498cd	3296 \pm 352a	135 \pm 11a	31 \pm 3b	121 \pm 3de	73 \pm 1abcd	30 \pm 1cde	58 \pm 3de	nd	nd	9177 \pm 855b	0.794 \pm 0.075b
Jiefangzhong	6964 \pm 510b	1401 \pm 279g	55 \pm 5c	18 \pm 2c	95 \pm 8e	42 \pm 6d	10 \pm 3de	177 \pm 29cd	1 \pm 0d	8 \pm 0b	8773 \pm 778b	0.736 \pm 0.101bc
Fuyang	4909 \pm 365de	2163 \pm 156bcd	72 \pm 5bc	10 \pm 1ef	222 \pm 22abc	80 \pm 10abcd	17 \pm 2de	215 \pm 52bc	nd	nd	7688 \pm 272c	0.692 \pm 0.066bcd
Moriowase	4140 \pm 238ef	2538 \pm 125bc	42 \pm 7c	16 \pm 2cd	277 \pm 32a	85 \pm 11abc	20 \pm 3de	2 \pm 1e	nd	nd	7120 \pm 176cd	0.659 \pm 0.070cde
Huobao 2	4797 \pm 349de	2115 \pm 103bcde	29 \pm 6c	11 \pm 1ef	211 \pm 2abcd	49 \pm 9cd	12 \pm 3de	4 \pm 1e	nd	nd	7228 \pm 394cd	0.633 \pm 0.057cdef
Xiangzhong 10	4576 \pm 324de	2335 \pm 326bed	30 \pm 1c	12 \pm 0de	247 \pm 12ab	51 \pm 2cd	59 \pm 4bc	355 \pm 38b	nd	nd	7665 \pm 380c	0.628 \pm 0.064cdef
Baihua	4149 \pm 385ef	1982 \pm 107cdef	62 \pm 8bc	12 \pm 2de	261 \pm 11a	72 \pm 13abcd	17 \pm 1de	232 \pm 17bc	nd	nd	6787 \pm 445cde	0.613 \pm 0.055cdef
Toi	3577 \pm 133fg	1997 \pm 273cdef	44 \pm 7c	10 \pm 1ef	259 \pm 12a	74 \pm 11abcd	38 \pm 14cde	159 \pm 47cde	nd	nd	6158 \pm 293defg	0.613 \pm 0.046cdef
Duobao 2	3109 \pm 362gh	2339 \pm 243bcd	57 \pm 1c	20 \pm 1c	219 \pm 4abc	62 \pm 3bcd	56 \pm 2bc	32 \pm 2b	2 \pm 0c	4 \pm 0e	5900 \pm 608efg	0.599 \pm 0.043def
Wugongbai	2418 \pm 671h	2616 \pm 592b	73 \pm 5bc	10 \pm 0ef	257 \pm 34a	52 \pm 5cd	16 \pm 1de	994 \pm 191a	nd	nd	6436 \pm 905def	0.537 \pm 0.062ef
Luoyangchin	4370 \pm 466ef	1390 \pm 344g	27 \pm 5c	8 \pm 1ef	272 \pm 22a	54 \pm 1cd	36 \pm 14cd	2 \pm 0e	3 \pm 0b	nd	6162 \pm 176defg	0.507 \pm 0.061f
Mogi	4424 \pm 290ef	1460 \pm 166fg	31 \pm 1c	10 \pm 0ef	222 \pm 6abc	50 \pm 3cd	37 \pm 3cd	4 \pm 1e	1 \pm 0d	nd	6239 \pm 224defg	0.502 \pm 0.060f
Zaozhong 6	4156 \pm 833ef	1001 \pm 54g	31 \pm 5c	12 \pm 1de	157 \pm 12bcde	65 \pm 9bcd	15 \pm 2de	29 \pm 2de	nd	nd	5466 \pm 320fgh	0.440 \pm 0.0589f
Tantou 4	2518 \pm 706h	1725 \pm 29efg	23 \pm 4c	8 \pm 2ef	210 \pm 16abcd	110 \pm 16a	18 \pm 3de	2 \pm 0e	nd	nd	4614 \pm 741hi	0.330 \pm 0.056g
Baili	3438 \pm 86fg	1377 \pm 115g	70 \pm 2bc	13 \pm 0de	127 \pm 16cde	96 \pm 10ab	4 \pm 0e	129 \pm 81cde	1 \pm 0d	6 \pm 0d	5262 \pm 67gh	0.240 \pm 0.034gh
Changhong 3	1299 \pm 137i	2107 \pm 182bcd	27 \pm 1c	7 \pm 0f	102 \pm 7e	88 \pm 5abc	9 \pm 1de	54 \pm 5de	1 \pm 0d	1 \pm 0f	3695 \pm 422i	0.165 \pm 0.033h
Meam	4399	2042	54	15	210	71	33	186			7013	0.599
Max	8912	3296	135	38	277	110	126	994			12958	1.270
Min	1299	1001	23	7	95	42	04	Trace			3695	0.165
SD	1741	569	31	8	62	20	31	295			2080	0.243
CV (%)	39.6	279	56.9	55.8	29.4	27.9	92.2	158.8			29.7	40.6

n = 5; Data followed by different letters within the same column are significantly different by Duncan's multiple range test at 0.05 level; CV: Coefficient of variation; nd: not detected.

The most common measure of acidity is titratable acidity, since it correlates well with perceived sourness^[16]. Sensory analysis experiments with artificial acid solution showed that the perceived acidity closely correlated with titratable acidity, and to a less extent pH^[29-30]. As shown in Table 1, considerable variations in titratable acidity of ripe pulp exist between cultivars. Clustering analysis based on titratable acidity showed that all 18 cultivars were basically classified into 5 groups: very high acidity, high acidity, medium acidity, low acidity, and very low acidity. Very high acidity group only included ‘Xiaomaopipa’, with a titratable acidity of 1.270%. High acidity, medium acidity, low acidity, and very low acidity groups consisted of 4, 6, 4 and 3 cultivars, respectively, with a titratable acidity range of 0.692% ~ 0.818%, 0.599% ~ 0.659%, 0.440% ~ 0.537% and 0.165% ~ 0.330%, respectively (Table 1, Fig. 3).

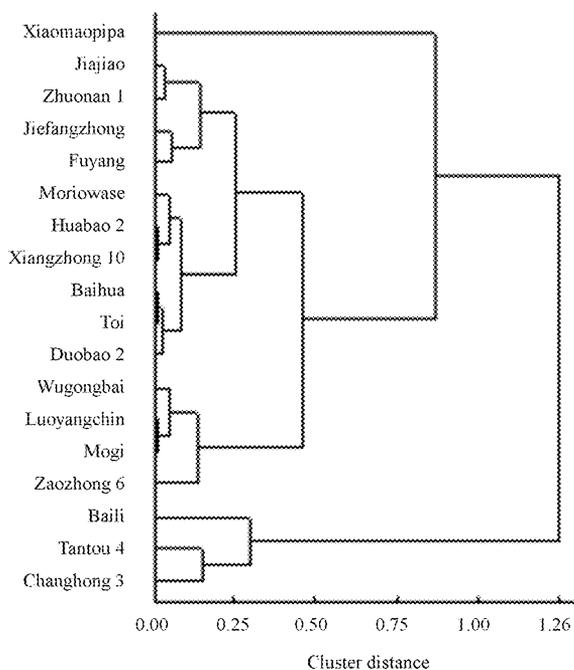


Fig. 3 Cluster analysis for 18 loquat cultivars based on titratable acidity of ripe pulp

Distribution of organic acids in the ripe fruit of 2 cultivars is shown in Table 3. In ‘Jiefangzhong’ and ‘Zaozhong 6’, no significant difference was found in

the absolute amounts of total acids and titratable acidity between pulp and skins, while the amounts of total acids and titratable acidity in seeds was much lower than in pulp and skins. Previous study has shown that grape skins have the highest total acid content, followed by pulp and seeds^[9]. This is basically consistent with our results. Like pulp, both skins and seeds of ‘Jiefangzhong’ and ‘Zaozhong 6’ presented 8 organic acids: malic, quinic, citric, iso-citric, α -ketoglutaric, fumaric, oxaloacetic and tartaric acids. Skins also contained trace quantities of ferulic, *cis*-aconitic and β -coumaric acids. Malic and quinic acids were the main acids in pulp, skins and seeds. In pulp and skins, content of malic acid was much higher than that of quinic acid, but in seeds, content of malic acid was slightly lower than that of quinic acid. The sum of malic acid plus quinic acid presented 86.9% ~ 88.6% of the total acids in skins and 94.3% ~ 95.5% in pulp; other acids attained only trace to 5.2% of the total acids in both pulp and skins. In seeds, the sum of malic acid plus quinic acid presented only 53.5% ~ 57.7% of the total acids; citric, α -ketoglutaric and tartaric acids accounted for 15.6% ~ 17.0%, 11.0% ~ 11.8% and 8.3% ~ 12.3% of the total acid content, respectively; other acids presented only 0.1% ~ 5.2%.

It is evident from this study that the composition of organic acids in loquat ripe pulp depends largely on cultivars. This work presents the first report of organic acids in loquat fruit tissues (pulp, skins and seeds). The wide range of diversity in malic acid content and titratable acidity in loquat ripe pulp among different cultivars may enable breeders to breed and select cultivars with improved flavor based on superior phenotypes. Knowledge of the organic acid composition of loquat fruit may be of use to food technologists and consumers.

Acknowledgement We thank Germplasm Repository for Loquat in Fujian Fruit Research Institute, Fujian Academy of Agricultural Sciences for providing loquat fruit.

Table 2 Correlation of matrix among organic acids of loquat ripe pulp

	Malic	Quinic	Citric	Iso-citric	α -Ketoglutaric	Fumaric	Oxalacetic	Tartaric	Total acids	Titrateable acidity
Malic	1.000									
Quinic	0.1264	1.0000								
Citric	0.4932*	0.5997**	1.0000							
Iso-citric	0.7265***	0.5146*	0.8124***	1.0000						
α -Ketoglutaric	0.0988	0.2170	-0.1310	-0.0611	1.0000					
Fumaric	-0.1028	0.1085	0.2303	0.2021	-0.0498	1.000				
Oxalacetic	0.6190**	0.3483	0.3379	0.6524**	0.4588	0.1118	1.000			
Tartaric	0.2573	0.3272	0.4705*	0.3464	0.2726	0.0394	0.3828	1.000		
Total acids	0.9310***	0.4493	0.6659**	0.8253***	0.2143	-0.0366	0.7063**	0.4701*	1.000	
Titrateable acidity	0.8782***	0.4823*	0.6021**	0.7902***	0.3689	-0.0672	0.7452***	0.4417	0.9644***	1.0000

*, **, ***: Significant at $P < 0.05$, 0.01 or 0.001 level, respectively.

Table 3 Organic acid composition (mg kg^{-1} FW) in pulp, skins and seeds from the ripe fruit of 2 loquat cultivars

Cultivars	Tissue	Malic	Quinic	Citric	Iso-citric	α -Ketoglutaric	Fumaric	Oxalacetic	Tartaric	Ferulic	<i>cis</i> -Aconitic	β -Coumaric	Total acids	Titrateable acidity (%)
Jiefangzhong	Pulp	6964 \pm 510a	1401 \pm 279b	55 \pm 5c	18 \pm 2c	95 \pm 8c	42 \pm 6c	10 \pm 3b	177 \pm 29b	1 \pm 0b	8 \pm 0a	2 \pm 0b	8773 \pm 778a	0.736 \pm 0.101a
	Skins	6079 \pm 42b	1520 \pm 123a	114 \pm 16b	49 \pm 3a	335 \pm 23a	159 \pm 9a	17 \pm 7a	455 \pm 13a	2 \pm 0a	7 \pm 0b	3 \pm 0a	8728 \pm 92a	0.710 \pm 0.021a
	Seeds	466 \pm 30c	784 \pm 48c	369 \pm 18a	39 \pm 3b	256 \pm 8b	63 \pm 2b	10 \pm 1b	179 \pm 9b	nd	nd	nd	2166 \pm 147b	0.220 \pm 0.019b
Zaozhong 6	Pulp	4156 \pm 833a	1001 \pm 54b	31 \pm 5c	12 \pm 1b	157 \pm 12c	65 \pm 9c	15 \pm 2c	29 \pm 2b	nd	nd	nd	5466 \pm 320a	0.440 \pm 0.059a
	Skins	3348 \pm 78b	1598 \pm 42a	78 \pm 2b	13 \pm 4b	299 \pm 15a	132 \pm 16a	25 \pm 4a	92 \pm 3a	Trace	Trace	Trace	5585 \pm 49a	0.433 \pm 0.018a
	Seeds	458 \pm 17c	529 \pm 18c	288 \pm 4a	40 \pm 4a	203 \pm 7b	95 \pm 4b	2 \pm 0b	227 \pm 15b	nd	nd	nd	1842 \pm 58b	0.148 \pm 0.010b

n = 5; Data followed by different letters within the same column for the same cultivar were significantly different by Duncan's multiple range test at 0.05 level; nd: not detected.

References

- [1] Cunha S C, Fernandes J O, Ferreira I M P L V O. HPLC/UV determination of organic acids in fruit juices and nectars [J]. *Eurp Food Res Techn*, 2002, 214: 67–71.
- [2] Koyuncu F. Organic acid composition of native black mulberry fruit [J]. *Chem Nat Comp*, 2004, 40: 367–369.
- [3] Lin S Q, Sharpe R H, Janick J. Loquat: botany and horticulture [J]. *Hort Rev*, 1999, 23: 233–276.
- [4] Chen F X, Liu X H, Lin H Y, et al. Determination of the organic acids from the fruit and leaf of loquat by ion-exchange chromatography [J]. *J Fujian Agri For Univ (Nat Sci)*, 2004, 33: 195–199. (in Chinese)
- [5] Ding C K, Chachin K, Ueda Y, et al. Metabolism of phenolic compounds during loquat fruit development [J]. *J Agri Food Chem*, 2001, 49: 2883–2888.
- [6] Kurssanow A L. Biochemistry of the ripening of fruits of the Japanses medlar (*Eriobotrya japonica*) [J]. *Planta*, 1932, 15: 752–766.
- [7] Rajput C B S, Singh J P. Chemical analysis of loquat fruits [J]. *Ind J Hort*, 1964, 21: 204–205.
- [8] Shaw P E, Wilson C W. Determination of organic acids and sugars in loquat (*Eriobotrya japonica* Lindl.) by high-pressure liquid chromatography [J]. *J Sci Food Agri*, 1981, 32: 1242–1246.
- [9] Lamikanra O, Inyang I D, Leong S. Distribution and effect of grape maturity on organic acid content of red muscadine grapes [J]. *J Agri Food Chem*, 1995, 43: 3026–3028.
- [10] MacRae E A, Bowen J H, Stec M G H. Maturation of kiwifruit (*Actinidia deliciosa* cv. Hayward) from two orchards: differences in composition of the tissue zones [J]. *J Sci Food Agri*, 1989, 47: 401–416.
- [11] Monselise S P, Galily D. Organic acids in grapefruit tissues [J]. *J Amer Soc Hort Sci*, 1979, 104: 895–897.
- [12] Kallio H, Hakala M. Sugars and acids of strawberry varieties [J]. *Eurp Food Res Techn*, 2000, 212: 81–85.
- [13] Randhawa G S, Singh R K N. The loquat in India [J]. *Ind Counc Agri Res Bull*, 1970, 24: 58.
- [14] Bentley R. The shikimate pathway — a metabolic tree with many branches [J]. *Crit Rev Biochem Mol Biol*, 1990, 25: 307–384.
- [15] Tiitinen K M, Yang B R, Haraldsson G G, et al. Fast analysis of sugars, fruit acids, and vitamin C in sea buckthorn (*Hippophae rhamnoides* L.) varieties [J]. *J Agri Food Chem*, 2006, 54: 2508–2513.
- [16] Lobit P, Soing P, Génard M, et al. Theoretical analysis of relationships between composition, pH, and titratable acidity of peach fruit [J]. *J Plant Nutr*, 2002, 25: 2775–2792.
- [17] Moing A, Svanella L, Rolin D, et al. Compositional changes during the fruit development of two peach cultivars differing in juice acidity [J]. *J Amer Soc Hort Sci*, 1998, 123: 770–775.
- [18] Souty M, Genard M, Reich M, et al. Effect of assimilate supply on peach fruit maturation and quality [J]. *Can J Plant Sci*, 1999, 79: 259–268.
- [19] Wu B H, Quilot B, Genard M, et al. Changes in sugar and organic acid concentrations during fruit maturation in peaches, *P. davidiana* and hybrids as analyzed by principal component analysis [J]. *Sci Hort*, 2005, 103: 429–439.
- [20] Drake S R, Eisele T A. Carbohydrate and acid contents of Gala apples and Bartlett pears from regular and controlled atmosphere storage [J]. *J Agri Food Chem*, 1999, 47: 3181–3184.
- [21] Drake S R, Eisele T A, Elfving D C, et al. Effects of the bioregulators aminoethoxyvinylglycine and ethephon on brix, carbohydrate, acid, and mineral concentrations in ‘Scarletspur Delicious’ apple juice [J]. *HortScience*, 2005, 40: 1421–1424.
- [22] Ethlenfeldt M K, Meredith F I, Ballington J R. Quince organic acid profile of rabbiteye vs. highbush blueberries [J]. *HortScience*, 1994, 29: 321–323.
- [23] Ayaz F A, Kadioglu A, Acar C, et al. Effect of fruit maturation on sugar and organic acid composition in two blueberries (*Vaccinium acetostaphylos* and *V. myrtilus*) native to Turkey [J]. *New Zeal J Crop Hort Sci*, 2001, 29: 137–141.
- [24] Marsh K B, Richardson A C, MacRae E A. Early- and mid-season temperature effects on the growth and composition of satsuma mandarins [J]. *J Hort Sci Biotechn*, 1999, 74: 443–451.
- [25] Cheng C H, Seal A G, Bolding H L, et al. Inheritance of taste characters and fruit size and number in a diploid *Actinidia chinensis* (kiwifruit) population [J]. *Euphytica*, 2004, 138: 185–195.
- [26] Marsh K, Attanayake S, Walker S, et al. Acidity and taste in kiwifruit [J]. *Postharvest Biol Technol*, 2004, 32: 159–168.
- [27] Okuse I, Ryugo K. Compositional changes in the developing ‘Hayward’ kiwi fruit in California [J]. *J Amer Soc Hort Sci*, 1981, 106: 73–76.
- [28] Silva B M, Andrade P B, Mendes G C, et al. Study of the organic acids composition of quince (*Cydonia oblonga* Miller) fruit and jam [J]. *J Agri Food Chem*, 2002, 50: 2313–2317.
- [29] CoSeteng M Y, McLellan M R, Downing D L. Influence of titratable acidity and pH on intensity of sourness of citric, malic, tartaric, lactic and acetic acids solutions and on the overall acceptability of imitation apple juice [J]. *Can Inst Food Sci Technol J*, 1989, 1: 46–51.
- [30] Pangborn R M. Relative taste intensities of selected sugars and organic acids [J]. *J Food Sci*, 1963, 27: 726–733.